

Netherlands Food and Consumer Product Safety Authority Ministry of Economic Affairs



Comparison of NGS pipelines and traditional diagnostics in annual Daucus carota surveys

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Overview presentation

- Phytosanitary survey in carrot
- Traditional diagnostics
- Next Generation Sequencing approach
 - > Selection of reference genes
 - > Reference based detection
 - > De novo and blast based detection
- Comparison of costs and hands-on time



Phytosanitary survey Daucus carota (carrot)

- Annual survey in carrot since 2011
- Presence of
 - > Ca. Liberibacter solanacearum (CaLsol) (EPPO A1)
 - > Ca. Phytoplasma solani (CaPsol) (EU II/A2)





Symptomatic material

- ~130 inspections resulting in ~30 samples
 - Presently, both pests have not been detected
- Sampling of symptomatic field-grown carrots
 - > Discolored leaves (red, yellow)
 - > Stunted growth
 - > Formation of side roots
- Symptoms not specific to both pests
 - Ca P. asteris suspected causal agent in majority of cases







Diagnostic testing scheme

- Two subsamples (leaves & carrot)
- Detection by:
 - > Conventional PCR CaPsol (leaves)
 - > Real-time CaLsol (leaves & carrot)
- Verification for selected samples
 - > targeted PCR Sanger sequencing

The iterative use of test methods:1. is time consuming2. requires lots of hands-on time3. is therefore costly





Next Generation Sequencing (NGS); an alternative?

- Why using the *Daucus carota* survey?
 - > Specific scope: 2 pests in symptomatic material (analytical sensitivity)
 - > Availability of reference sequences (analytical specificity)
 - > Survey shared by multiple disciplines
 - * "Long" turn-over time allowed



Analysis pipelines – reference vs. de novo





Defining suitable reference sequences

Entire genome not suitable for detection

- CaPsol, CaLsol and CaPast genomes share homology (non-specific mapping)
- Regions with variable resolution (nonspecies level resolution)
- Determining cut-offs for detection using the entire genome is not possible



CaPsol sequence data mapped to CaPsol, CaPast and CaLsol



Selection of reference genes

Suitable reference genes are:

- Single copy orthologs (SCO)
 - > Even coverage expected
 - > Can be compared over species
- SCO with species level resolution
- SCO >500 nt for reliable mapping



107 reference genes per species



Not all 107 species specific SCOs can be used for RNAseq pipeline

- Selected reference genes for DNA pipeline are not equally transcribed
- Transcription level per SCO is conserved over samples
 - Highly expressed gene in sample 1 = highly expressed gene in sample 2
- 51 species specific SCOs with at least >5x average coverage in individual samples were selected for RNAseq pipeline





Reference assembly results – DNA and RNA

LIMS number	Traditional diagnostics		DNA NextSeq			RNA NextSeq		
	CaLsol/CaPsol detected	Suspected causal agent	CaLsol	CaPsol	CaPast	CaLsol	CaPsol	CaPast
5727954	NO	putative CaPast	NO	NO	+	NO	NO	+
5727962	NO	CaPast	NO	NO	+	NO	NO	+
5727970	NO	CaPast	NO	NO	+	NO	NO	+
5727989	NO	putative CaPast	NO	NO	+	NO	NO	+
5727997	NO	putative CaPast	NO	NO	+	NO	NO	+
5728009	NO	putative CaPast	NO	NO	+	NO	NO	+
5728228	NO	putative CaPast	NO	NO	+	NO	NO	+
5728244	NO	CaPast	NO	NO	+	NO	NO	+
5728367	NO	unknown	NO	NO	NO	NO	NO	NO
5728375	NO	CaPast	NO	NO	+	NO	NO	+
5728391	NO	CaPast	NO	NO	+	NO	NO	+
5728420	NO	putative CaPast	NO	NO	+	NO	NO	+
5728439	NO	putative CaPast	NO	NO	+	NO	NO	+
5728447	NO	unknown	NO	NO	NO	NO	NO	NO
5728455	NO	putative CaPast	NO	NO	+	NO	NO	+
5728463	NO	unknown	NO	NO	NO	NO	NO	NO
5728471	NO	CaPast	NO	NO	+	NO	NO	+
5974066	NO	putative CaPast	NO	NO	+	NO	NO	+
5974074	NO	unknown	NO	NO	NO	NO	NO	NO
6792299	NO	unknown	NO	NO	NO	NO	NO	NO

- Identical qualitative results were obtained from the DNA and RNA detection pipelines
- Pipeline output could easily be interpreted



de novo assembly + blast-based detection

- Beyond the initial scope of the survey
- When the usual suspects cannot be detected, are there other possible causal agents that could explain the symptoms observed?
- Blast-based detection: indicative and use with caution!
- Interactive visualisation tool: Krona





Possible candidates for observed symptoms

• Carrot viruses detected:

Carrot torrado virus 1

- Carrot cryptic virus
- Carrot mottle virus

Carrot red leaf luteovirus associated RNA

Carrot read leaf virus

 Carrot read leaf virus and Carrot mottle virus were detected in all CaPast negative samples (#5) and possibly causing the observed symptoms of Carrot motley dwarf (CMD)



What about the money?

- Direct costs are higher per sample
- Hands-on time is greatly reduced per sample
 - > Traditional: 57 min/sample
 - > NGS: 21 min/sample
- Net extra costs per sample: €89
 - > Saving in hands-on time
 - > Re-usable datasets
 - Possibilities for detection beyond initial scope of survey

Traditional diagnostics	CO	costs per		
	sa	mple		
DNA extraction (Qiagen, plant mini kit)	€	8,00		
Bio-X-ACT short mix reactions	€	2,00		
TaqMan universal Master Mix	€	9,00		
Qiaquick PCR Purification Kit	€	2,00		
BigDye terminator kit v1.1	€	21,00		
DyeEx kit	€	6,00		
Pop7 polymer	€	1,00		
Capillary	€	2,00		
Total	€	51,00		

NGS approach (DNA)	costs per sample	
DNA extraction (Qiagen, plant mini kit)	€	4,00
NGS NextSeq, DNA, PE150, 4.5 Gb data	•	320,00
Total	€	324,00



Conclusions

• We created a robust and reliable detection pipeline for CaPsol, CaLsol and CaPast detection in symptomatic carrot material





Future work

- Create scripts for automated generation of result forms including results and conclusions for the different analyses
 - > User-friendly, interactive, but stand-alone and write protected (QA)
 - > In close collaboration with specialists from different disciplines
- Determine performance criteria following PM7/98(2) for the analysis pipelines and compare those to traditional tests
- Increase computational power and storage for NGS data



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Thank you for your attention





RNAseq is not properly mapped to the host

de Novo assembly and blast-based detection

Sample_01 (CaPast positive)

