
– EPPO Workshop –

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Breeding for insect resistance in OSR: is it a dream?

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Insect-resistant cultivars in other species



Constraints

Very few major resistance genes

→ Need for (semi-)quantitative phenotyping

→ Labor intensive, slow process



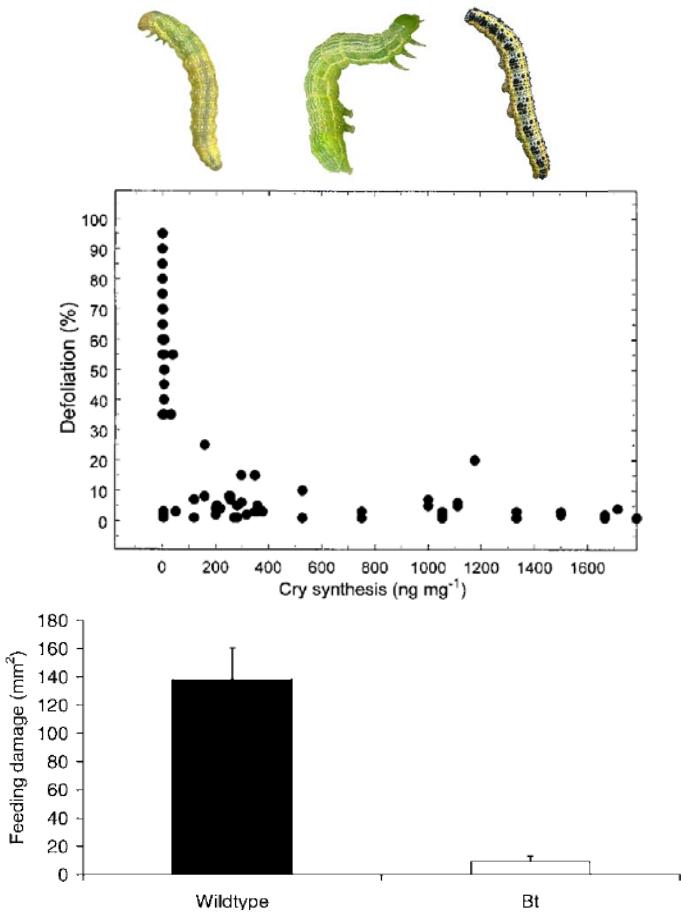
→ Insect availability



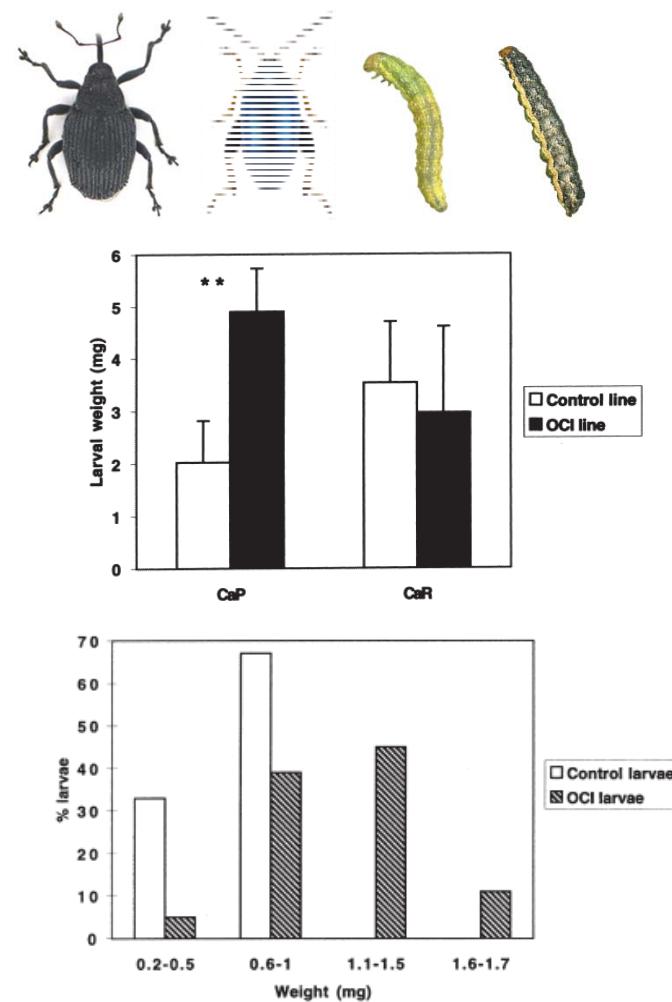
BOTTLENECK



Strategy 1: transgenes

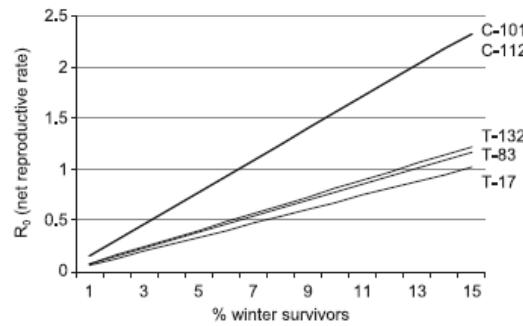
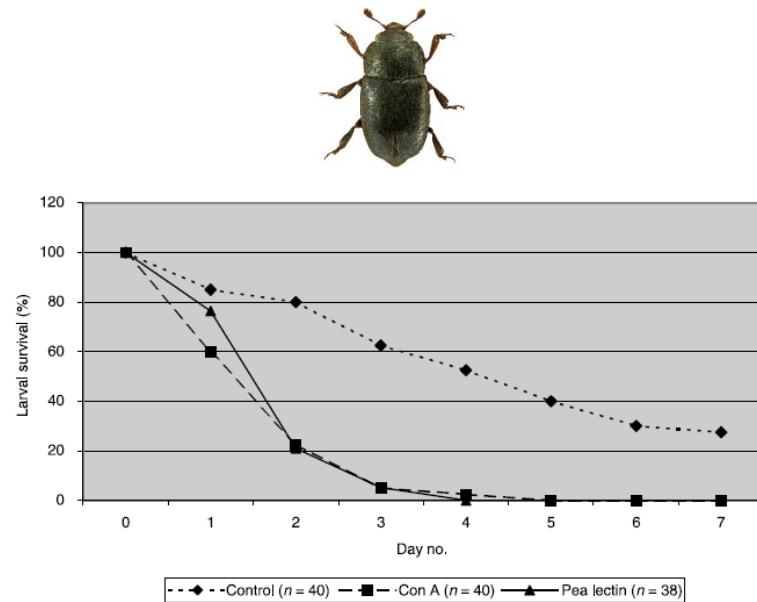


Stewart *et al.* 1996, 1997, Ramachandran *et al.* 1998a,b, Halfhill *et al.* 2001, 2002, Mason *et al.* 2003, Sayyed *et al.* 2003, Schuler *et al.* 2003, 2004, Cerdà *et al.* 2006, Le *et al.* 2007, Walker *et al.* 2007, Himanen *et al.* 2008, 2009, Ibrahim *et al.* 2008, Kjaer *et al.* 2010

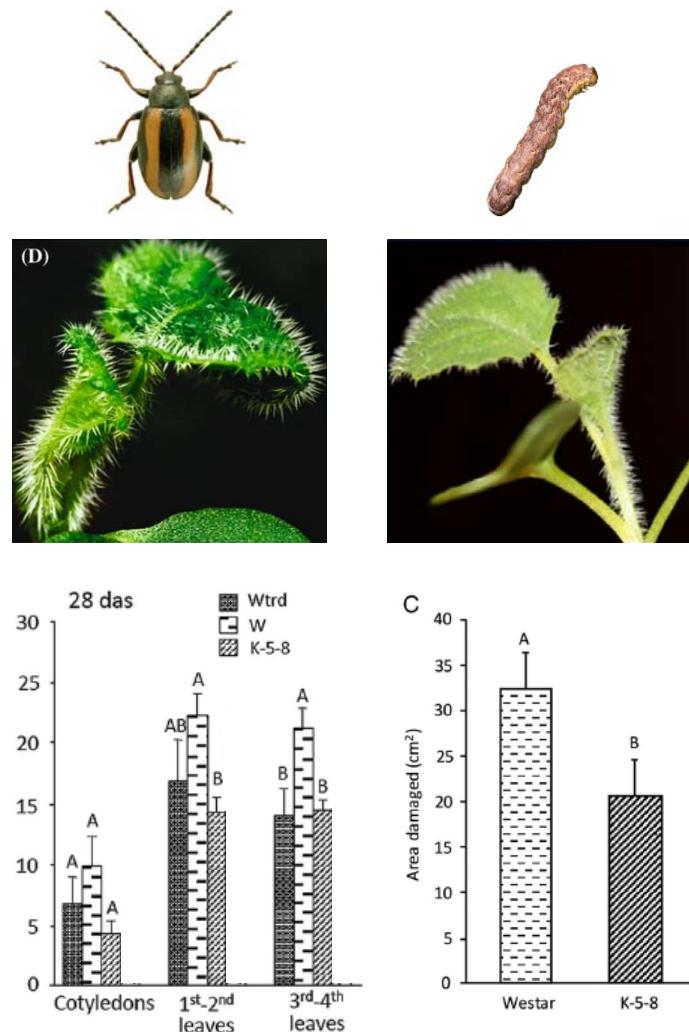


Bonadé-Bottino 1993, Girard *et al.* 1998a,b, Bonadé-Bottino *et al.* 1999, De Leo *et al.* 2001, Ferry *et al.* 2005

Strategy 1: transgenes

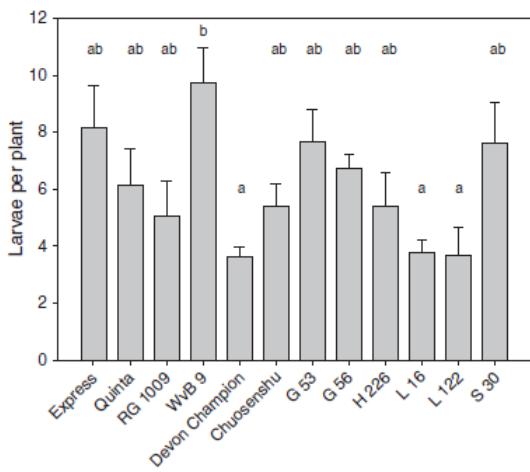


Åhman & Melander 2003, Melander *et al.* 2003,
Åhman *et al.* 2006, Lehrman 2007,
Lehrman *et al.* 2007, Åhman *et al.* 2009

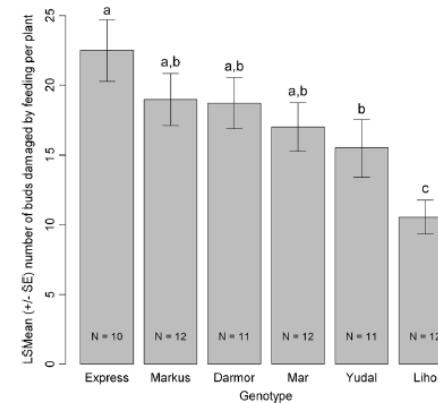


Gruber *et al.* 2006, Soroka *et al.* 2011,
Alahakoon *et al.* 2016a,b

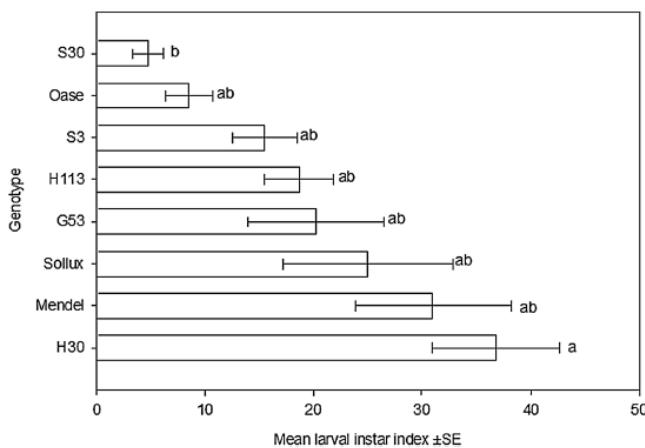
Strategy 2: natural resistance



Eickerman & Ulber 2010, Eickerman *et al.* 2011



Åhman 1993, Charpentier 1986, Hervé *et al.* 2014a,b, 2016a,b



Schäfer-Kösterke *et al.* 2017, Schäfer *et al.* 2017

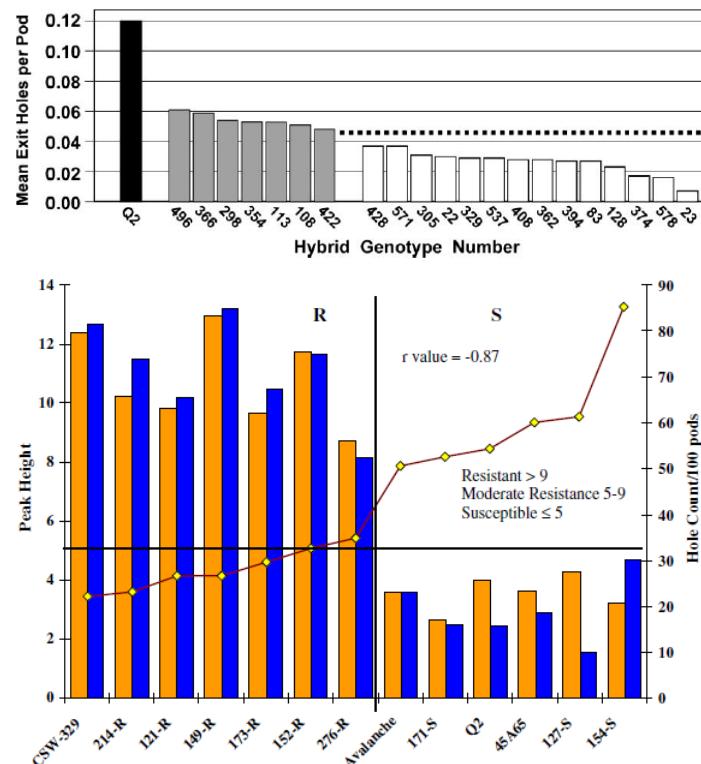


NOTHING

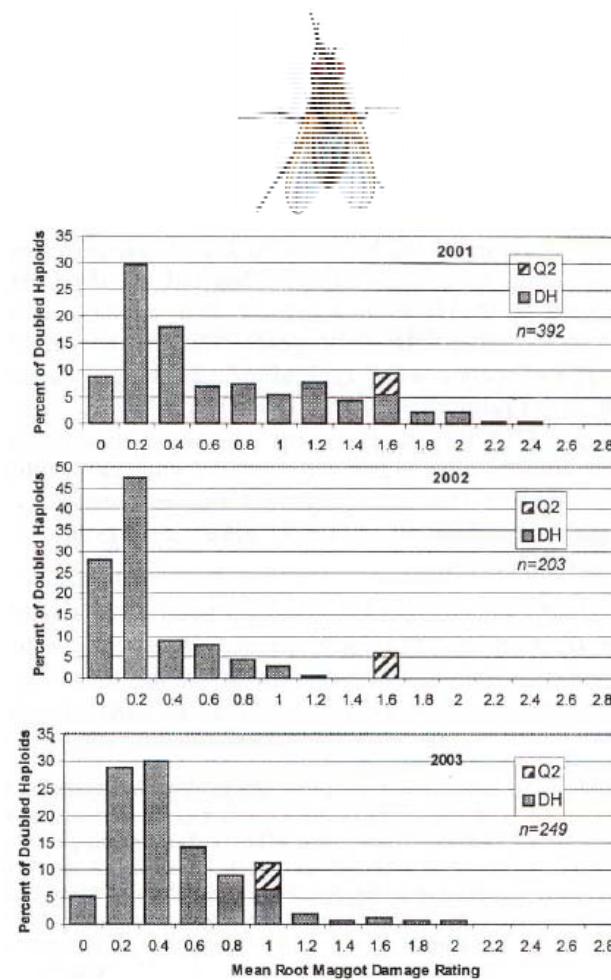
Dosdall *et al.* 1994, unpublished results



Strategy 3: introgression from other *Brassicaceae*

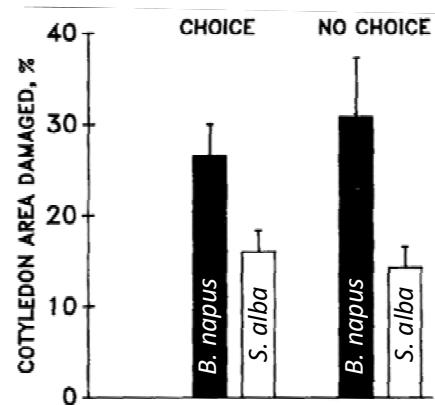


Doucette 1947, Brown *et al.* 1999, Kalischuk & Dosdall 2004, McCaffrey *et al.* 2004, Dosdall & Kott 2006, Ulmer & Dosdall 2006, Cárcamo *et al.* 2007, Shaw *et al.* 2009, Tansey *et al.* 2010a,b,c, Lee *et al.* 2014



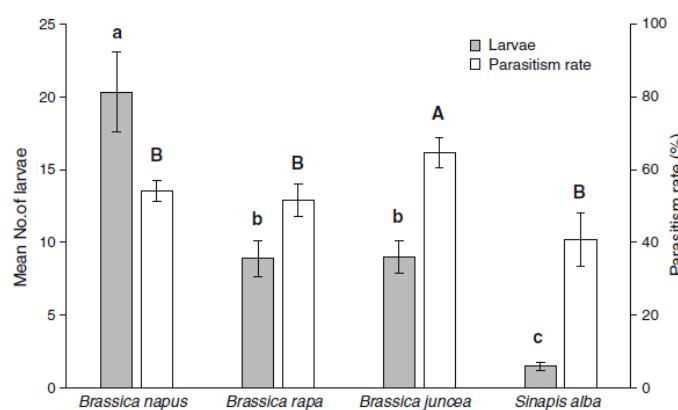
Dosdall *et al.* 1994, 2000, Kott & Dosdall 2004, Ekuere *et al.* 2005

Strategy 3: introgression from other *Brassicaceae*



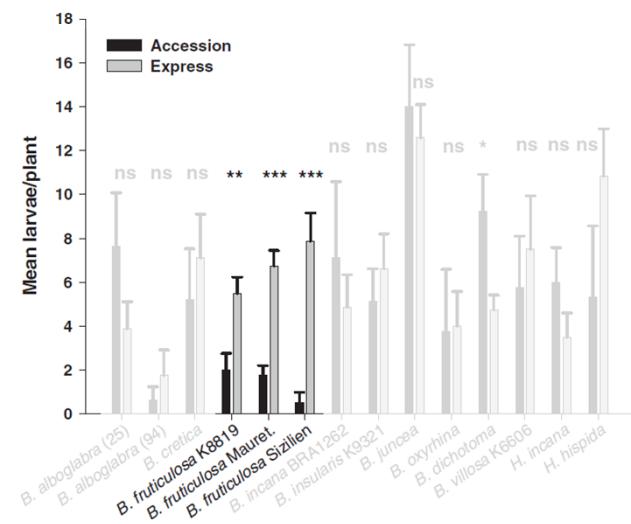
Species	Line	Larval weight (mg)	% larvae reaching 4th instar	% survival	Damage rating
Experiment 1					
<i>S. alba</i>	AC Pennant	28.9±4.2bcd	48±8.5c	90±5.4	9.4
	L-GS	23.6±1.2cd	48±6.7c	94±6.0	9.1
<i>B. carinata</i>	Dodolla	37.4±3.1abc	54±8.4bc	86±8.0	6.9
	S-67	35.8±3.9abc	62±9.0abc	90±6.1	7.3
<i>B. juncea</i>	AC Vulcan	18.8±1.6d	2±2.0d	96±4.0	6.4
<i>B. rapa</i>	Echo	44.2±5.8a	56±11.0abc	80±10.0	7.5
	AC Boreal	43.7±3.0a	78±5.4a	98±2.0	9.1
<i>B. napus</i>	AC Excel	49.2±7.7a	68±10.8abc	86±6.1	7.8
	Midas	39.4±5.1ab	74±9.8ab	98±2.1	7.6
Experiment 2					
<i>B. juncea</i>	AC Vulcan	24.6±5.8c	3±2.5c	83±4.4	4.1
	H-Allyl	26.5±6.5c	15±5.0c	83±4.5	4.6
	H-Butenyl	36.0±6.0bc	18±5.8bc	70±9.2	4.9
	L-GS	52.0±8.5ab	38±9.4ab	83±7.8	7.6
<i>B. napus</i>	AC Excel	56.3±7.2a	50±9.2a	95±3.2	8.3

Putnam 1977, Lamb 1980, 1984, Bodnaryk & Lamb 1991, Gavloski *et al.* 2000, Soroka & Grenkow 2013



Borg & Ekbom 1996, Ekbom & Borg 1996, Hopkins & Ekbom 1996, 1999, Ekbom 1998, Hopkins *et al.* 1998, Veromann *et al.* 2012, 2014, Kaasik *et al.* 2014a,b

Ulmer *et al.* 2001, 2002, Dosdall & Ulmer 2004



Eickerman & Ulber 2010

Perspectives

Most transgenic OSR are based on the production of toxins

- qualitative strategy
- same problems as with insecticides

Quantitative resistance is likely to be more durable...

... but it will probably not be possible based on existing material

- introgression from other brassicaceous species (*S. alba*)
- resynthesized OSR

Challenge of phenotyping → (bio)marker-assisted selection

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Thank you for your attention





Flea beetles (*Phyllotreta* sp.)



Cabbage stem flea beetle
(*Psylliodes chrysocephala*)



Rape winter stem weevil
(*Ceutorhynchus picitarsis*)



Rape stem weevil
(*Ceutorhynchus napi*)



Cabbage stem weevil
(*Ceutorhynchus pallidatylus*)
(*= C. quadridens*)



Cabbage seed weevil
(*Ceutorhynchus obstrictus*)
(*= C. assimilis*)



Pollen beetle
(*Meligethes aeneus*)
(*= Brassicogethes aeneus*)



Cabbage root fly (= root maggot)
(*Delia radicum*)



Brassica pod midge
(*Dasineura brassicae*)



Turnip sawfly
(*Athalia rosae*)



Bertha armyworm
(*Mamestra configurata*)



Cabbage aphid
(*Brevicoryne brassicae*)



Green peach aphid
(*Myzus persicae*)